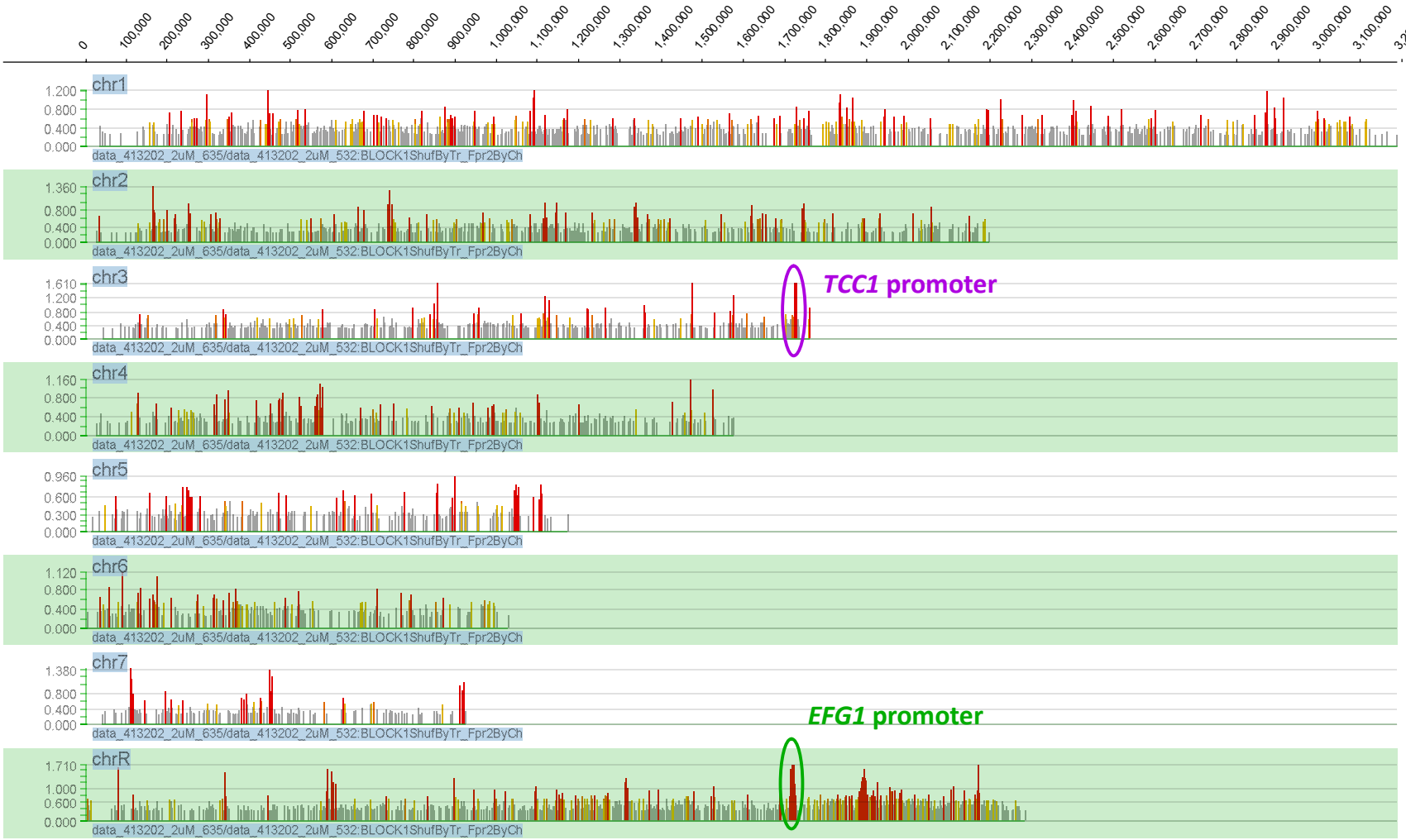

Figure S1. Binding of Efg1 to chromosomal sequences of the *C. albicans* yeast growth form.
Strain HLCEEFG1 was grown in YPD medium at 30 °C in the yeast form and chromosomally bound HA-Efg1 was fixed with formaldehyde. Fragmented chromatin was immunoprecipitated with anti-HA antibody, labelled and used as probe for microarrays covering the *C. albicans* genome. Untagged strain DSC11 was used to determine background values for each region. Regions significantly binding to HA-Efg1 are colored according significance ratings, red lines indicate the most significant and grey lines the least significant binding regions.

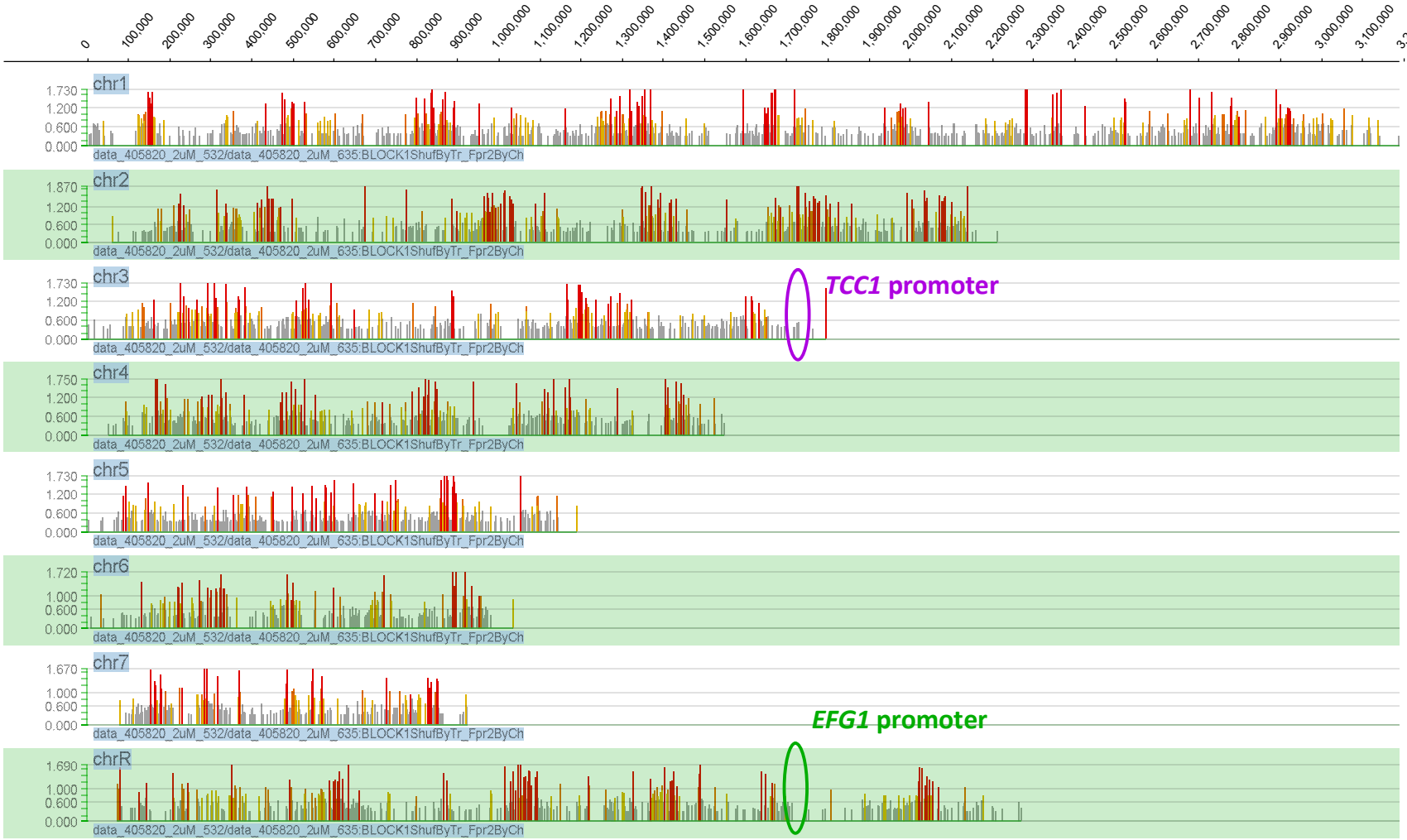

Figure S2. Binding of Efg1 to chromosomal sequences of *C. albicans* after hyphal induction.
Strain HLCEEFG1 was pregrown in YPD medium at 30 °C and hyphae were induced in YP medium containing 10 % serum for 30 min at 37 °C. Cells were harvested and chromosomally bound HA-Efg1 was fixed with formaldehyde. Fragmented chromatin was immunoprecipitated with anti-HA antibody, labelled and used as probe for microarrays covering the *C. albicans* genome. Strain DSC11 producing untagged Efg1 was used to determine background values for each region. Regions significantly binding to HA-Efg1 are colored according significance ratings, red lines indicate the most significant and grey lines the least significant binding regions.

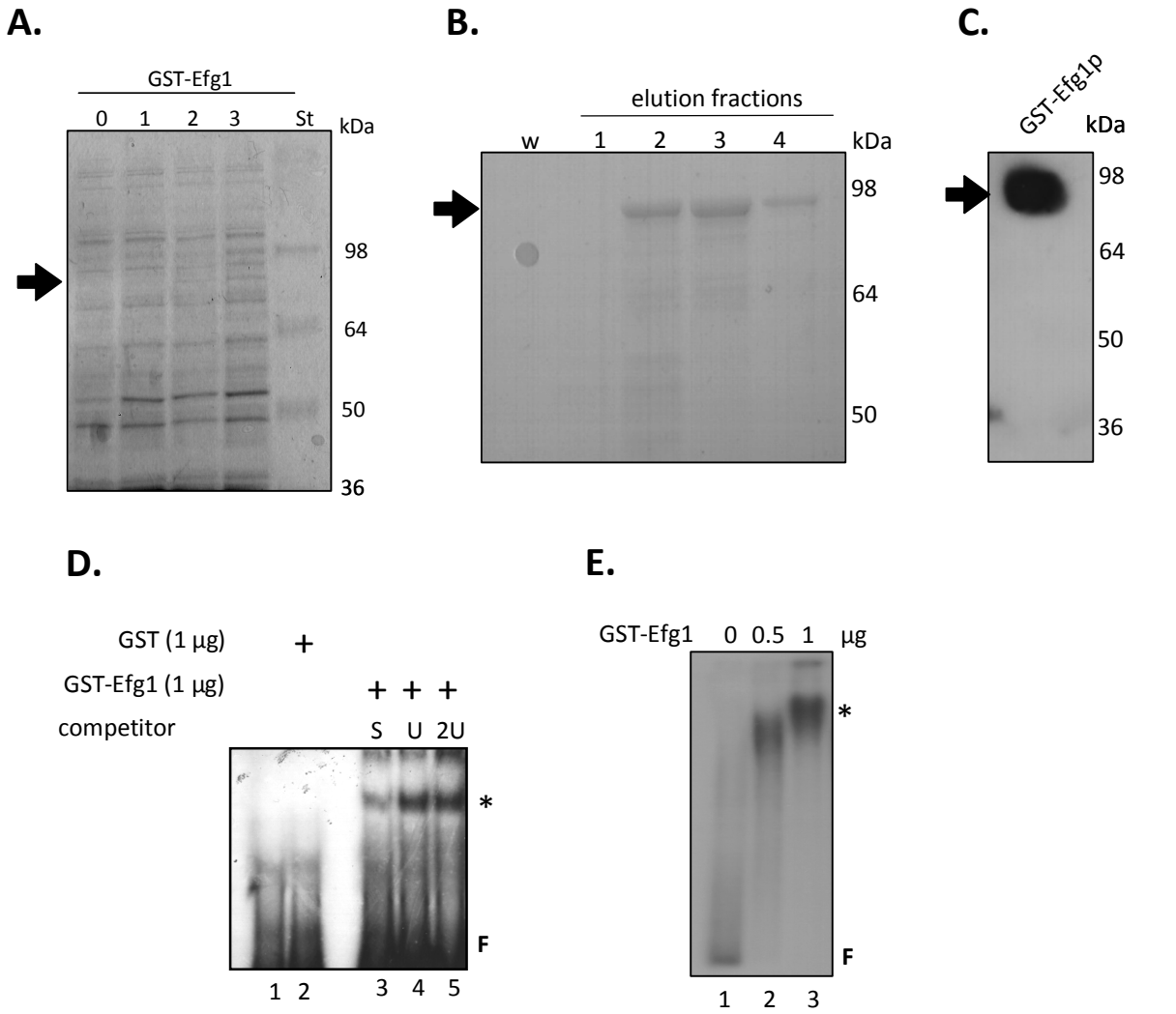

Figure S3. Purification and DNA binding of GST-Efg1. (A) *E. coli* strain BL21 transformants carrying expression vector pMi-11 were grown in LB medium at 37 °C to an $OD_{600\ nm}$ of 0.8; then 0.5 mM IPTG was added and incubation was continued for 3 h at 30 °C. Cells were analysed by SDS-PAGE before (t=0) and 1, 2 and 3 h after IPTG-addition and proteins were visualised with Coomassie Brilliant Blue. St, standard proteins; the arrow indicates the migration of GST-Efg1. (B) Cells were induced with IPTG for 3 h and then loaded on a glutathione-Sepharose 4B column. The column was washed with PBS and then eluted with glutathione-containing elution buffer. Samples of the wash (w) and subsequent elution fractions 1-4 were separated by SDS-PAGE and stained by Coomassie Brilliant Blue. (C) An aliquot of elution fraction 2 was separated by SDS-PAGE and then analysed by immunoblottings, using anti-GST monoclonal antibody as primary and anti-mouse antibody as secondary antibody.

Binding of GST-Efg1 to MCB-containing oligonucleotide (P1/P2-3xMCB) (D) and 408 bp *EFG1* promoter fragment (E). Both DNAs were ^{32}P-labelled and GST-Efg1 protein was used in the indicated concentrations for EMSA assays. Binding reactions contained 1 µg (lane 4) or 2 µg (lane 5) poly(dI-dC) as unspecific competitors (U) or a 5fold excess of unlabelled oligonucleotide as specific competitor (S). Positions of DNA-protein complexes and non-bound DNA are indicated, respectively, by asterisks and "F".